AL-FARABI KAZAKH NATIONAL UNIVERSITY

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WORKING TOGETHER

AN EXAMPLE OF SUCCESSFUL SCIENTIFIC Collaboration between Pakistan AND KAZAKHSTAN



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The Center of Physical and Chemical Methods of Research and Analysis





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RESEARCH PROJECTS PAK-KAZAKH SCIENTIFIC COLLABORATION PROJECT ENTITLED, "STUDIES on the BIOLOGICALLY ACTIVE METABOLITES from the PLANTS of CENTRAL ASIA"

N. A. Sultanova, M. Iqbal Choudhary, T. Makhmoor, V. B. Omurkamzinova, Atta-ur-Rahman, and Z. A. Abilov Chemical investigation of genus Tamarix - T. hispida, T. ramosissima 24 A.F. Miftahova, V. U. Ahmad, G. S. Burasheva, and Z. A. Abilov Biological Active Compounds from Halostachys caspica, Halocnenum F. Miftahova, A. Dar, A. U.Vikar, G. S. Burasheva, and Z. A. Abilov Bioassay of Extracts from Halocnemum strobilaceum, Suaeda Physophora, L. Korulkina, G. E. Zhusupova, Z. A. Abilov and M. Iabal Choudhary R. A. Muzichkina, Y. A. Litvinenko, M. Iqbal Choudhary and T. Makhmoor Method of Obtaining of Polyphenol Complex with Antioxidant Activity..... 55 B. K. Yeskaliyeva, G. S. Burasheva., M. Iqbal. Choudhary and Z. A. Abilov A. K.Umbetova, N. A. Sultanova, V.B. Omurkamzinova, M. Igbal Choudhary and Z. A. Abilov Chemical Research of Kazakhstan euhalophyte species C. monspeliacum of Camphorosma genus and T. laxa, T. elongata of Tamarix genus of Z. Z. Karzhaubekova., B. S. Siddiqui, G. S. Burasheva and N. A. Sultanova

Chemical Research of Kazakhstan euhalophyte species C. monspeliacum of Camphorosma genus and T. laxa, T. elongata of Tamarix genus of Chenopodiaceae, Tamaricaceae families

A. K.Umbetova, N. A. Sultanova, V.B. Omurkamzinova, M. Iqbal Choudhary and Z. A. Abilov

The objects of our investigation are the over-ground mass of Camphorosma monspeliacum (Chenopodiaceae family), Tamarix laxa and Tamarix elongata (Tamaricaceae family) plants. They were sampled in Almaty and Aral (dried bottom) areas in the blooming phase.

Isolation and Separation of Biologically Active Compounds

For extraction of bioactive substances from C. monspeliacum, T. laxa and T. elongate, comparative research of propyl, ethyl and methyl alcohols, acetone and their aqueous solutions was carried out. It was determined that the greatest BAS quantity is extracted by 70%-ethyl alcohol during 72 hours at room temperature. The obtained wateralcohol extracts were filtered then and concentrated in the vacuum of water jet pump until the full alcohol was removed. Prior the separation of bioactive substances, the fractional extraction of water-alcohol extracts of C. monspeliacum, T. laxa and T. elongata plants was carried out by means of chloroform and ethyl acetate. Chloroform, ethyl acetate and water solutions of investigated plant species were analyzed by means of GLC, PC and TLC methods. Chloroform extracts of C. monspeliacum, T. laxa and T. elongata plants contain chlorophylls, lipophilic substances, high saturated and unsaturated carboxylic acids (fatty acids) and terpenoids. However, the extracts of C. monspeliacum contain chromones. Ethyl acetate extracts of C. monspeliacum, T. laxa and T. elongata plants contain phenolic acids, flavonoids and their glycoside forms. Water solutions contain aminoacids and carbohydrates. Additionally, sulphate forms of flavonoids and hydrolysable tanning agents were identified in the water solutions of T. laxa and T. elongata plants. Hence, the 20 substances were shown in the water-alcohol extracts of *C. monspeliacum* plants and 30 substances were obtained in the water-alcohol extracts of *T. laxa* and *T. elongata* plants.

Extraction of individual plants was carried out by means of adsorption distributing chromatography (polyamide, silica gel), gel chromatography (LH-20), preparative HPLC, PC and TLC. 10 individual compounds from *C. monspeliacum* plants: 3 terpenoids (substances 2.1, 2.4 and 2.5), 3 chromones (substances 2.6, 2.7 and 2.8), 3 flavonoids (substances 2.14, 2.20 and 2.22) and 1 phenolic acid (substance 2.17).

26 individual compounds were extracted from *T. laxa* and *T. elongata* plants: 3 terpenoids (substances 2.1 - 2.3). 17 Flavonoids (substances 2.9-2.16, 2.19-2.25, 2.26-2.28), 4 phenolic acids (substances 2.17, 2.18, 2.29, 2.30) and 2 hydrolyzable tannins (substances 2.31, 2.32) were also extracted.

Determination of Terpenoids Structure

Ain accordance to the results of TLC (the developer is sulfate of cerium) and positive reaction with the Liberman's reagent it is stated that the main components of chloroform extract are terpenoids: substances 2.4 and 2.5 from the plants of *C.monspeliacum*, substances 2.1-2.3 are from the plants of *T. laxa* and *T. elongata*. Five substances (2.1 – 2.5) were evolved from the chloroform extracts of *T. laxa*, *T. elongata* and *C. monspeliacum* by employing adsorption distributing chromatography (silica gel) and preparative TLC techniques.

Substance 2.1 was obtained from three investigated species of plants; substances 2.2 and 2.3 were obtained from the plants of *T. laxa* and *T. elongata* and substances 2.4 and 2.5 were obtained from the plants of *C. monspeliacum*.

On the basis of physical-chemical data and in comparison with the literature data substance **2.1** is β -sitosterol; substance **2.2** is methyl ether of the 3- β -al-D-fridoolean-14-en-28 carboxylic acid; substance **2.3** is 3- α -[3",4"-Dihydroxy-trance-cinnamyl-oxy-D-fridoolean-14-en-28-carboxy-lic acid (izotamarixen); substance **2.4** is 3-O- β -D-glucopiranoside of the oleanolic acid; substance **2.5** is 28-O- β -D-glucopiranoside of the oleanolic acid.

Earlier β -sitosterol was identified by the others foreign and home scientists for the plants of *Chenopodiaceae* and *Tamaricaceae* families. It 64

should be noted that β -sitosterol was found in all the investigated plants. Substance **2.3** was earlier obtained from Kazakhstan species *Tamarix hispida*. Substances **2.4** and **2.5** were described in literature, but for the first time they were obtained from the plants of *Camphorosma* genus.

Substance **2.2** (0.34%) was related to pentacycle triterpenoids because of claret coloring by sulfate of cerium and Liberman-Burhard reaction. In IR spectrum there are absorption bands which are typical for methyl, methyne, methylene and ketonic groups in the range of 2864-2933 cm⁻¹ and 1689 cm⁻¹, respectively. In the mass spectrum the peak of molecular ion (*m*/*z* 482) corresponding to C₃₂H₅₀O₃ molecular formula was registered by means of EI-MS and FAB-MS (-ve) methods. Fragments with m/*z* 248, 204, 189 and 133 correspond to retro-dien disintegration on Diels-Alder, which is typical for pentacycle triterpenoids (Fig. 3).

Also, 32 signals of the carbon atoms were observed in a spectrum of NMR ¹³C. By means of DEPT in a normal phase seven methyl (C-23-27, C-29-30), four methyne (C-5, C-9, C-15,C-18), and in a return phase ten methylene carbon atoms (C-1, C-2, C-6, C-7, C-11, C-12, C-16, C-19, C-21, C-22) were registered. Proton signals of seven methyl groups in a spectrum NMR ¹H were registered at δ 0.82-0.98 as 3H singlets, and twenty protons of methylene groups were registered as multiple signals at δ 1.08-1.98.

Besides, in a spectrum NMR ¹H at δ 5.54 olefinic proton resounds at double bond H-15 (1H, dd, J₁ 11.0 and J₂ =3.4 Hz,), indicating that substance **2.2** refers to taraxeran-14-en type or to D-fridoolean derivatives, and at δ 3.30 to the β -form.

In ¹³C NMR spectrum, ¹³C signals of carbon atoms with double bond (C-14 and C-15) resound at δ 161.0 and δ 117.0; signals C-28 and C-3 resound at δ 179.0 and δ 76.0, respectively. Singlet at δ 3.80 in a spectrum of NMR of ¹H and the area δ 56.0 in a spectrum of NMR ¹³C is typical for the methyl ether of carbonyl group at C-28, but in a spectrum NMR ¹³C of the substances **2.2** an additional signal of aldehydic carbon atom was found out at δ 238.0 The presence of carbonyl groups of aldehydic and methyl ether was also proved by means of massspectrometry, fragment with m/z 438 (Fig. 1).

COSY-45" and HMBC spectroscopy were used to prove the double bond position of COOCH₃, CHO and CH₃ groups. Carbon atom C-15 in 65 the HMBC spectrum interacts with the protons H-16, H-18, and C-14 with the protons at C-26, C-18. The COSY-45° spectrum also confirms the interaction of H-15 and H-16 protons. Hence, according to 2 D spectra the double bond is located between C-14 and C-15 atoms.

Position of COOCH₃ group was proved by the HMBC spectrum: signals of protons H-18, H-16 and H-22 interact with carbon atom at C-28 and C-17 positions. Carbon atom of aldehydic group interacts with a methyl proton at C-23 (δ 0.98), that confirms addition of the aldehydic groups at C-3 position. Besides, in the NMR ¹H spectrum the doublet of doublets signal with J₁ = 9.0, J₂ = 4.7 Hz, indicates the fact that the methyn proton at C-3 is in an axial, and aldehydic group is in an equatorial position. The methyl groups' positions were completely proved to be true by the HMBC spectrum. On the basis of physical and chemical analysis methods the structure for substance 2.2 was stated as methyl ether of the 3- β -al-D-fridoolean-14-en-28 carboxylic acid. The **substance 2.2 is a new chemical compound not described in the literature earlier**.



Figure 1 – Fragmentation scheme of methyl ether 3-β-al-D-fridoolean-14-en-28 carboxylic acid (2.2)

Chemical study of the genus *Camphorosma* plants (*C. monspeliacum*) and the genus *Tamarix* plants (*T. laxa, T. elongata*) indicates the prescence of pentacylic tri terpernoid. Such conclusion was made on the basis of study of the individual substances, by means of chroma-

tographic and spectral analysis methods. Thus, olean triterpenoids are typical for the plants of Camphorosma genus (C. monspeliacum) and fridoolean triterpenoids are typical and for the plants of Tamarix genus (T. laxa and T. elongata).

Determination of Structure Chromones

This class of natural chemical compounds was evolved from the chloroform extract of C. monspeliacum on silica gel and elucated, eluating with hexane and hexane-acetone mixture. Substances 2.6 (0.44%) and 2.7 (0.02%) gave bluish-dark blue fluorescence in UV-light and yellow coloring in ammonia vapor and with sulfate of cerium. In the UV spectrum of substances 2.6 and 2.7 in area $\lambda_{max} = 207-283$ and 306-368 nm two absorption bands were observed and the absorption band of carbonyl group in the range of 1654 cm⁻¹ (2.6) and 1641 cm⁻¹ (2.7) in the IR spectra, are typical for chromones, that is accounted for low negative inductive influence of the γ -pyrone cycle hetero atom. For substance 2.7 in the range of 3456 cm⁻¹ the absorption band of hydroxyl group was additionally developed.

Molecular masses of substances 2.6 and 2.7 (by means of HREIMS method) are 268 (for substance 2.6) and 314 (for substance 2.7), that correspond to formulae $C_{16}H_{12}O_4$ (for substance 2.6) and $C_{17}H_{14}O_6$ (for substance 2.7). Besides, in the NMR ¹³C spectrum (methods BB, DEPT) sixteen carbon atoms were registered for substance 2.6 and seventeen carbon atoms were registered for substance 2.7.

Substances 2.6 and 2.7 are primarily related to the derivatives of 2phenoxychromone. In mass-spectrum the formation of the chromone nucleus with m/z 148 was ac companied with the formation of the phenoxyl with m/z 92. The presence of the characteristic fragment with m/z 120 corresponds to retrodiene disintegration mechanism of ypyronic cycle of chromone nucleus. In ¹³C NMR spectrum the the C-2 and C-1' atom signals within the limits at δ 166.0-169.0 and δ 144.0-148.0, accordingly were typical for 2- phenoxychromones' system. In the spectrum NMR ¹³C of substances 2.6 and 2.7the C-2 signals resound at δ 166.5 (2.6) and δ 166.6 (2.7); and the C-1' signals resound at δ 148.0 (2.6) and δ 147.5 (2.7), that corresponds to the literature data.

In the spectrum of NMR 1H, substances 2.6 and 2.7 the proton signals at C-3 are at δ 6.84 (substance 2.6) and δ 6.71 (substance 2.7) 67 intervals as one-proton singlet, and carbon atoms at C-3 in the spectrum of NMR ¹³C are shown at δ 116.2 (substance **2.6**) and δ 108.9 (substance **2.7**), respectively. Two multiple signals in spectrum NMR ¹³C of substance **2.6** at δ 8.03 (2H, H-2 ', H-6 ') and δ 8.04-8.06 (3H, H-3 ', H-4 ', H-5 ') specify on the unreplaced phenoxyl ring. For substance **2.7** signals of the phenoxyl protons were shown at δ 7.56 (2H, d, J =7.9 Hz, H-3 ', H-5 ') and δ 7.97 (2H, d, J =8.0 Hz, H-2 ', H-6 '), that confirms 4 '-replacement.

In the spectrum of NMR ¹H, of substance **2.6** the proton of the chromone benzol ring at C-5 was exposed to disscreening influence of carbonyl group at C-4 and as distinct from the other protons exposed in weaker area at δ 7.58 (1H, d, J = 7.0 Hz). The signals H-6 (1H, dd, J₁ which were 8.8, J₂ = 2.2 Hz) and H-8 (1H, d, J 2.1 Hz) were shown at δ 7.09 and δ 7.23.

In substance 2.7 the benzene ring protons of chromone nucleus were shown at δ 6.54 and δ 6.81 areas as doublets with intensity in 1H with 1.9 and 2.0 Hz belonging to meta-interacting protons H-6, H-8; and H-6 signal resounds in a stronger field than H-8.

The location of chromone benzene ring signals in spectrum NMR ¹³C of substance 2.7, also, corresponds to 5,7- dire placement. The C-6 signals were found at δ 97.5, and the C-8 signals were found at δ 94.3.

Besides in spectra NMR ¹H and in NMR ¹³C of substances **2.6** and **2.7**, signals belonging to -OCH₃ group were registered. In mass-spectrum (methods EI / MS, FAB-MS (-ve) the presence of one -CH₃ group in substance **2.6** and two -CH₃ groups in substance **2.7** were confirmed by observed characteristic fragments with m/z 252 (**2.6**) and 285 (**2.7**). The presence of one -OH group in substance **2.7** was proved to be true by formation of the fragment with m/z 251.

Determination of a fine structure of substances was proved to be true by means of 2D correlation HMBC spectra and hence proton at C-3 of pyrone cycle interacts with carbon atoms C-2, C-3 and C-4, respectively. The -OCH₃ groups' position was also proved to be true by means of HMBC spectrum. Three – proton singlet of the proton of -OCH₃ group of substance **2.6** interacts with the carbon atom in the C-7 position; and in substance **2.7** the interaction of two -OCH₃ groups' atoms with C-5 and C-7 carbon atoms was marked. On the basis of spectral and chemical analysis methods, the structure of the substances 2.25 was confirmed by the acid hydrolysis data and by the presence of optical rotation.

On the basis of physical and chemical characteristics, spectral analysis methods and according to the literature data, substance 2.12 was identified as 5,7,4 '-Trihydroxy-3'-methoxyflavon (chrysoeriol); substance 13 was identified as 7,3',4'- Trihydroxy-5-methoxyflavon.

For substances **2.9**, **2.10**, **2.11**, **2.14**, **2.15** and **2.16**, the following structures were identified: 3,7,3',4'-Tetrahydroxy-5-methoxyflavon (2.9); 3,5,7-trihydroxy-3',4'-dimethoxyflavon (2.10); 3,5,4'- trihydroxy-7,3'-dimethoxyflavon (ramnazine) (2.11); 3,5,7,3',4'-Pentahydroxyflavon (quercetin) (2.16); 3,5,7,4'-Tetrahydroxy-3'-methoxyflavon (isorhamnetin) (2.14); 3,5,7,3'-tetrahydroxy-4'-methoxyflavon (tamarixetine) (2.15).

Substance **2.19** was characterized as 3-O- β -D-glucopyranoside of kaempferide; **2.25** – 3-O- β -D-glucopyranoside of quercetin; **2.22** – 3-O- β -D-glucopyranoside of isorhamnetine; **2.20** – 3-O- β -D-galactopyranoside of isorhamnetine; **2.23** – 3-O- β -D-glucopyranoside of tamarixe-tine; **2.21** – 3-O- α -L-rhamnopyranoside of tamarixetine; **2.24** – 3-O- α -L-rhamnopyranoside of tamarixetine; **2.24** – 3-O- α -L-rhamnopyranoside of tamarixetine;

Substances **2.14** and **2.22** were obtained from all the investigated species of plants (*C. monspeliacum*, *T. laxa* and *T. elongata*). For the plants of *Camphorosma* genus the substances **2.14** and **2.22** were new and for the plants of *Tamarix* genus substance **2.22** was also obtained for the first time. Substance **2.21** was new for the plants of *Camphorosma* genus, and substances **2.12**, **2.13**, **2.9-2.11**, **2.19**, **2.23** and **2.24** were new for the plants of *Tamarix* genus.

The chemical composition of the investigated species of euhalophyte plants of *Camphorosma* genus (*C. monspeliacum*) and *Tamarix* genus (*T. laxa, T. elongata*) showed, that flavonoids basically were presented in the form of methoxylazed flavonol (**2.9, 2.10, 2.11, 2.14** and **2.15**). The glycosidized forms were also found (**2.19-2.24**). The glycosidation in flavonols is carried out on OH-group at the C-3 aglycon position. The basic flavonol sugar component is glucose (**2.19, 2.22** and **2.23**). However, there were flavonols carbohydrate, part of which was presented by rhamnose (**2.21**), arabinose (**2.24**) and galactose (**2.20**). It should be noted, that flavonols (substances **2.12** and **2.13**) were obtained from the ethyl-acetate extract of the *T. laxa* and *T. elongata* plants, as distinct from the *C. monspeliacum* plants.

2.25 was confirmed by the acid hydrolysis data and by the presence of optical rotation.

On the basis of physical and chemical characteristics, spectral analysis methods and according to the literature data, substance 2.12 was identified as 5,7,4 '-Trihydroxy-3'-methoxyflavon (chrysoeriol); substance 13 was identified as 7,3',4'- Trihydroxy-5-methoxyflavon.

For substances 2.9, 2.10, 2.11, 2.14, 2.15 and 2.16, the following structures were identified: 3,7,3',4'-Tetrahydroxy-5-methoxyflavon (2.9); 3,5,7-trihydroxy-3',4'-dimethoxyflavon (2.10); 3,5,4'- trihydroxy-7,3'-dimethoxyflavon (ramnazine) (2.11); 3,5,7,3',4'-Pentahydroxyflavon (quercetin) (2.16); 3,5,7,4'-Tetrahydroxy-3'-methoxyflavon (isorhamnetin) (2.14); 3,5,7,3'-tetrahydroxy-4'-methoxyflavon (tamarixetine) (2.15).

Substance **2.19** was characterized as 3-O- β -D-glucopyranoside of kaempferide; **2.25** – 3-O- β -D-glucopyranoside of quercetin; **2.22** – 3-O- β -D-glucopyranoside of isorhamnetine; **2.20** – 3-O- β -D-galactopyranoside of isorhamnetine; **2.23** – 3-O- β -D-glucopyranoside of tamarixetine; **2.21** – 3-O- α -L-rhamnopyranoside of tamarixetine; **2.24** – 3-O- α -L-arabopyranoside of tamarixetin.

Substances **2.14** and **2.22** were obtained from all the investigated species of plants (*C. monspeliacum*, *T. laxa* and *T. elongata*). For the plants of *Camphorosma* genus the substances **2.14** and **2.22** were new and for the plants of *Tamarix* genus substance **2.22** was also obtained for the first time. Substance **2.21** was new for the plants of *Camphorosma* genus, and substances **2.12**, **2.13**, **2.9-2.11**, **2.19**, **2.23** and **2.24** were new for the plants of *Tamarix* genus.

The chemical composition of the investigated species of euhalophyte plants of *Camphorosma* genus (*C. monspeliacum*) and *Tamarix* genus (*T. laxa*, *T. elongata*) showed, that flavonoids basically were presented in the form of methoxylazed flavonol (**2.9**, **2.10**, **2.11**, **2.14** and **2.15**). The glycosidized forms were also found (**2.19-2.24**). The glycosidation in flavonols is carried out on OH-group at the C-3 aglycon position. The basic flavonol sugar component is glucose (**2.19**, **2.22** and **2.23**). However, there were flavonols carbohydrate, part of which was presented by rhamnose (**2.21**), arabinose (**2.24**) and galactose (**2.20**). It should be noted, that flavonols (substances **2.12** and **2.13**) were obtained from the ethyl-acetate extract of the *T. laxa* and *T. elongata* plants, as distinct from the *C. monspeliacum* plants.

Determination of the obtained Structure of Hydrolysable Tannins

The water extracts of the *T. laxa* and *T. elongata* plants containing several substances which are primarily related to hydrolyzable tannins were determined by means of the two-dimensional PC method with the specific developers' application. From the water extract two individual substances **2.31** and **2.32** were obtained by means of adsorption-distributive and gel-chromatography methods. On the basis of chemical (acid hydrolysis) and physical and chemical analysis methods in comparison with the literature data substance **2.31** had 2-galloyl-D-glucopyranose and compound **2.32** had 3-O-(1-dehydrodigalloyl)-4,6-hexahydroxydiphenoylglucopyranose structure. Compound **2.32** was obtained as a result of acid hydrolysis from *T. aphylla* and from Kazakhstan species *T. hispida*.

Thus, such individual substances as vanillic acid, isorhamnetin, 3-Oβ-D-glucopyranoside of isorhamnetine, β-sitosterol and glucose were obtained from all the investigated euhalophyte plants of *Camphorosma* genus (*C. monspeliacum*) and *Tamarix* genus (*T. laxa*, *T. elongata*).

The genus *Camphorosma* (*C. monspeliacum*) are characterized with the triterpenoids of the oleanoic type and 2-phenoxychromomes; plants of *Tamarix* genus (*T. laxa* and *T. elongata*) are characterized with triterpenoids of the fridooleanoic type, tanning agents and sulphate form of flavonoids.

Methyl ether of 3-β-al-D-fridoolean-14-en-28 carboxylic acid obtined from *T. laxa* and *T. elongata* (substance **2.2**) and *C. monspeliacum* – 7-methoxy-2-phenoxychromone (**2.6**), 5,7-dimethoxy-2-(4'-hydroxyphenoxy) chromone (**2.7**) are new natural compounds.

Biological Activities of Camphorosma genus Plants (C. monspeliacum) and Tamarix genus plants (T.laxa, T. elongata)

For studying the biological activities of Kazakhstan euhalophyte species *C. monspeliacum* of *Chenopodiaceae* family and *T. laxa* and *T. elongata* of *Tamaricaceae* family the water-alcohol, chloroform, ethyl acetate, water extracts, flavonoid fractions and individual substances **2.3, 2.6, 2.7, 2.11, 2.25** and **2.26** were tested by means of standard informative tests (*in vitro*) in specialized laboratories of the H.E.J Research Institute of Chemistry and in the specialized laboratories of Dr. Panjwani Center of Molecular Medicine and Drug Research, Karachi University (Pakistan). Water-alcohol, chloroform, ethyl acetate, water extracts from *C. monspeliacum*, *T. laxa* and *T elongata* possess antibacterial, antifungal and growth-regulating activities. The extracts from *T. laxa* and *T. elongata* have a high antioxidant activity. The extracts from *T. elongata* have a high antiamnezia activity.

Individual substances 2.3, 2.11 and 2.26 also have potential antioxidant and antidiabetic – activities.

For the first time toxicity of plant extracts of *Camphorosma* genus (*C. monspeliacum*) and *Tamarix* genus (*T. laxa* and *T. elongata*) was investigated against larvae of *Artemia salina* shrimp (Brine Shrimp activity) and their immunomodulatory activity was also performed. At the concentration of 0.4 mg / mL, the chloroform extract of *C. monspeliacum* (CM-4), containing substances **2.6** (KM – 1) and **2.7** (KM – 2) as basic components shows a high immunomodulatory activity. Water-alcohol extracted TE-1 of *T. elongata* containing the substance **2.25** (GE-19) also showed a high immunomodulatory activity. Individual substances **2.6** (KM – 1) and **2.7** (KM – 2) at the 0.4 mg / mL concentration also possess immunomodulatory activity, and the substance **2.25** (GE-19) in similar concentration (0.4 mg / mL) enhances its action.

For the first time immunomodulatory and *Brine Shrimp* activities (investigation of toxicity of vegetative extracts and biologically active substances by means of larvae of *Artemia salina*) were established for these euhalophyte species such as *C. monspeliacum*, *T. laxa* and *T. elongata*. Besides, immunomodulatory activity for class of chromones (compounds **2.6** and **2.7**) was also investigated for the first time.

Conclusion:

The following conclusions were made from results of research:

[1]. New sources of bioactive substances of plants of *Chenopodiaceae* family of *Camphorosma* genus (*C. monspeliacum*) and *Tamaricaceae* family of *Tamarix* genus (*T. laxa and T. elongata*) were revealed: chromones and triterpenoids of olean series were revealed in *C. monspeliacum*; triterpenoids of friloolean series, hydrolysable tannins and sulphate form of flavonoids were revealed in *T. laxa* and *T. elongata*; 12 fatty acids and 18 amino acids were shown by GLC; the mineral composition of ash residues were also determined.

[2]. The scheme of separation of BAS was developed: 10 individual substances were deduced from *C. monspeliacum*, they are 3 terpenoids, 3 chromones, 3 flavonoids, 1 phenolic acid; 26 individual substances were deduced from *T. laxa* and *T. elongata*, they were 3 terpenoids, 17 flavonoids, 4 phenolic acids and 2 hydrolyzable tannins.

[3]. For the first time three new compounds were isolated from *T*. *laxa* and *T*. *elongata* methyl ether of 3β-al-D-fridoolean-14-en-28carbonic acid; from *C*. *monspeliacum* 2-phenoxychromones – 7-methoxy-2-phenoxychromones and 5,7-dimethoxy-2-(4'-hydroxyphenoxy) chromone. 6 substances were new for the plants of *Camphorosma* genus and 11 substances were new for the *Tamarix* genus. Substances' structure was determined by means of chemical and spectral analysis methods (IR, UV, NMR ¹³C -, NMR ¹H-, 2D – COSY 45°, HNQC, HMBC, mass spectrometry).

[4]. 15 extracts and 6 individual substances were produced. Antibacterial, antifungal, antioxidant, antiamnezia, antidiabetic and growthregulating activities were shown. For the first time immunomodulatory and toxilogical activities (Brine Shrimp activity) were studied for the plants of *Camphorosma* genus (*C. monspeliacum*) and *Tamarix* genus (*T. laxa and T. elongata*). Immunomodulatory activity of chromones was also studied.